



Patient-Derived Tumour Organoids for Precision Screening: Operationalising GxP-Level Quality for Translational Reliability

There is a persistent gap between preclinical promise and clinical reality in oncology drug development. Two-dimensional cell culture systems derived from animal models cannot reproduce the architectural and microenvironmental complexity that shapes therapeutic responses in human tumours. Poor clinical outcomes despite encouraging preclinical data have prompted regulatory agencies to emphasise human-relevant test systems, with reduced reliance on animal models to improve translational accuracy.

Patient-derived organoids have emerged as an alternative that replicates the three-dimensional structure, genetic diversity and heterogeneity of human tumours. However, the translational utility of these organoids depends on two inseparable pillars: biological fidelity and operational rigor. Biological fidelity ensures that in vitro responses reflect patient outcomes, while operational rigor guarantees that results are reproducible and scalable across studies and client programmes. To implement organoids from exploratory tools to standardised, decision-grade platforms requires a quality management system that is GxP-aligned and optimised for precision screening, data integration and development.

Integrating Organoids into Development and Manufacturing Workflows

In the traditional workflow, organoid-based evaluation of drug efficacy ends when a contract development and manufacturing organisation (CDMO) is selected to produce first-in-human (FIH) material. In the proposed model, organoid screening and CDMO capabilities co-exist, ensuring a seamless transition from data-driven candidate selection to commercial manufacturing.

Housing both functions within one organisation has several advantages. Results from organoid testing directly inform CDMO involvement, creating a cohesive, evidence-based handoff that strengthens early development. The integration ensures that the screening stage considers manufacturability and that the assay design considers process feasibility, formulation constraints and expression, thereby preventing technically impractical candidates from advancing.

By governing discovery and manufacturing processes, the single GxP-aligned quality system ensures reliable documentation, deviation management and data compliance, which enhances traceability and aligns with regulatory expectations. Assay parameters, growth kinetics and analytical metrics are version-controlled to streamline knowledge transfer and preserve the scientific context related to critical quality attributes and processes.

The resulting smooth transition from post-evaluation to development increases the speed to market and eliminates ambiguity.

From tissue acquisition through assay readout to manufacturing, a robust chain-of-custody ensures a unified record of the origin and analytical outcome of each model. In addition, change-control and corrective-preventive action processes manage any deviations from the model that might have downstream impacts. Operators thus consistently reverify the results when they modify any key reagents, materials, or methods.

The unified governance structure ensures that human-relevant efficacy data and manufacturability criteria serve as the basis of portfolio decisions. As a result, drug developers can prioritise the programmes that are most feasible technically and likely to succeed clinically. Ultimately, using a single quality and operational framework when conducting organoid efficacy improves data continuity, reduces late-stage rework and establishes a scientifically grounded foundation for FIH readiness.

Genomic and Transcriptomic Validation to Ensure Biological Fidelity

The principal biological advantage of organoids lies in their capacity to reproduce tumour complexity in vitro. Their three-dimensional structure preserves cell-cell and cell-matrix interactions, which determine drug penetration, target engagement and downstream signaling. These elements are lost in two-dimensional monolayer cultures. While animal models provide in vivo contexts, they often fail to replicate human pharmacology or immune biology, limiting their predictive power. Regulatory roadmaps for New Approach Methodologies (NAMs) highlight organoids as credible, human-relevant systems when supported by validated and quality-managed workflows. Platforms need to combine biological fidelity with GxP-level quality controls to generate the data needed and ensure high-quality outputs for regulatory approval.

Analytical rigor is a prerequisite for achieving the clinical validity and regulatory acceptance of organoid-derived data in translational contexts. Multi-omic validation of organoid models verifies biological fidelity: Whole-exome sequencing (WES) confirms that organoids retain key driver mutations and co-alterations present in the source tumour, while RNA sequencing (RNA-seq) characterises transcriptional states and pathway activities influencing the drug response. To prevent the distortion of translational relevance caused by clonal selection or culture artifacts, organoids exceeding the predefined stability thresholds are retired. Each assay is validated for dynamic range, signal-to-background ratio and intra- and inter-plate precision. Dose-response curves are fitted against predefined acceptance criteria, including R^2 thresholds and Hill slope boundaries, while reference compounds, blinded duplicates and inter-site replicates quantify reproducibility.

The proposed platform integrates WES and RNA-seq data with matched clinical and pathological datasets, enabling the accurate prediction of response and resistance. Non-responding organoids



serve as the means to identify compensatory mutations or pathway activations that undermine the primary drug mechanism. Mapping these multi-omic features to phenotypic endpoints, such as viability, apoptosis, or image-based morphology, develops composite biomarker panels that move beyond single-gene predictors. A version-controlled, quality-managed repository curates all data, creating a foundation for future machine learning (ML) models to link the genotype, phenotype and clinical context to predictive outputs. A continuous bridge between discovery and clinical translation, formed through stratified libraries, tests against models representing specific patient populations by mechanism, pathway, or chemical series.

Operationally, organoid-based screening increases portfolio efficiency by reducing false positives, which are typical in oversimplified assays. Because screening occurs in a human-tumour-relevant architecture, hits are more likely to represent true target-driven activity, equipping drug developers with the capability to focus on candidates with greater translational potential. This approach produces leaner, stronger pipelines because fewer compounds advance, but those that do are supported by compelling evidence.

From Large-scale Screening to ML-enabled Prediction

The scale-up of organoid screening requires automation and robust data governance. Liquid-handling procedures, plating densities and assay timing must be version-controlled and environmental parameters, such as temperature and CO₂ stability, must be monitored and logged. Standardised imaging pipelines that use fixed exposure and segmentation settings are key and any software updates should trigger revalidation. These measures ensure consistency and a dataset that includes all the necessary information. All primary data files must be write-protected, timestamped and access-controlled to preserve traceability and integrity. These safeguards confirm that datasets are complete and suitable for cross-study comparison, meta-analysis and potential regulatory submission.

The FDA's roadmap for reducing animal testing underscores the importance of standardised, quality-managed organoid datasets in advancing human-relevant models. Although the current platform focuses on oncology, the same framework can be extended to other therapeutic areas once suitable tissues and endpoints are available. The GxP-aligned data architecture ensures the scale of the dataset, metadata uniformity and endpoint harmonisation, which ML depends on. Over time, validated predictive models that combine organoid phenotypes, genomic features and clinical contexts could guide trial design, dose selection and combination therapy strategies. As regulatory acceptance grows for NAMs, rigorously validated organoid data may increasingly supplement, or in some cases replace, animal data in early-stage drug development.

Bridging Current Gaps

Despite the promise of organoids, certain gaps remain. The success rate for organoid establishment varies by tumour type and time-to-assays may be a limiting factor for rapidly progressing cancers. Co-culture systems that incorporate immune or stromal components are under active development but are not yet standardised for large-scale application.

In addition, methodological heterogeneity between laboratories impedes cross-study data pooling. Addressing these challenges requires further standardisation, automation and multi-site validation under harmonised quality frameworks. Large prospective and harmonised studies will be essential to confirm clinical validity, demonstrate translational utility and confirm whether organoid-guided therapies improve patient outcomes when compared with standard care. These solvable challenges are contingent upon collaborative infrastructure and the adherence to shared quality standards.

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